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Taxonomic rank of Indian tortoise: revisit with DNA barcoding perspective

Abstract

Taxonomic rank of terrestrial tortoise inhabiting in India and adjacent countries has been an unresolved issue. Wild living species of Manouria and Indotestudo genera have been classified through conventional taxonomy, but not adequately discriminated by mitochondrial cytochrome b gene. Cytochrome oxidase (COI) marker has been quite successful to achieve the exact species level information. There needs of an accurate sequence based effort for the extant northeast (NE) Indian tortoises to identify them accurately and to provide locality information. To estimate the Kimura-2-Parameter (K2P) divergences and to construct the Neighbour-Joining (NJ) phylogeny, we generated six partial COI sequences derived from expert-identified tortoise specimens. Both BLASTn and Bold-IDs revealed the definitive identity of Manouria, whereas the congeners of Indotestudo remain inconclusive by only a two-fold interspecific divergence gap with the other named Indian tortoise species. The NJ phylogeny readily differentiated the three Indian tortoise genera with their respective conspecies but depicted a nonsynonomous group of Indotestudo congeners. This study shows that the molecular identification of Manouria emys phayrei, Indotestudo elongata and an allopatric population of Indotestudo forstenii in NE India corresponds correctly with existing morphological key support.

Keywords

Testudinidae • Northeast India • Morphology • Mitochondrial cytochrome oxidase (COI) • Distribution

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Introduction

Turtles and tortoises are the oldest living animals on the earth and play important roles in the food chains and life cycles within their environment [1,2]. Most threatened tortoise species occur in Asia [3-6]. India is reportedly inhabited by four tortoises, among them Indotestudo elongata and Manouria emys in the northeastern regions [7,8]. These land tortoises are the subject of many recurrent taxonomic controversies about their sporadic distributions. Also, all the land tortoises inhabiting in the Northeast (NE) India have been listed in Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES), Indian Wildlife (Protection) Act, 1972 (IWPA) and International Union for Conservation of Nature (IUCN), yet they are still poached [9], despite being the subject of lists e.g., CITES drawn up to protect wildlife against overexploitation and to prevent international trade from threatening species [10]. Therefore, it is a burning issue to protect the land tortoises of NE India before they reach the verge of extinction.

The Asian brown tortoise, *Manouria emys* is classified into two subspecies based on morphological key characters and distinct geographical distribution [11,12]. The other terrestrial elongated

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tortoise, *Indotestudo elongata* was selected as type species of the new subgenus *Indotestudo* [13]. Later on, cladistic analyses promoted *Indotestudo* to full generic rank, and it was reported as comprising of three species after re-examining the morphological key characters [14-16]. Furthermore, morphological as well as molecular data depict the presence of *Indotestudo elongata* in Arakan and Western Myanmar, *Indotestudo travancorica* in Western Ghats of South-Western India, and *Indotestudo forstenii* in Sulawesi and the adjacent island of Halmahera, so the species can be regarded as "Endemic in Indonesia" [17,18]. However, species identification and locality information obtained through the traditional taxonomy and limited molecular markers still appears intuitive.

Accurate species-level identification is strictly essential prior to the development of conservation application [19]. The identification helps to understand the species-specific life history and distribution patterns [20-22]. DNA barcoding of world's biodiversity has received momentum with the objective to enrich the Barcode of Life Data System (BOLD), a global reference library [23]. The barcoding technique utilizes a partial Cytochrome Oxidase c subunit I (COI) gene as a molecular marker that has been proven instrumental in identification at the species level

[24-28]. Distance based cladistic approach has furnished better resolution in species discrimination where the similarity search failed to identify the species correctly [29,30]. This study was intended to i) examine the taxonomic key character of *Manouria emys*, *Indotestudo elongata* and *Indotestudo forstenii* ii) analyze the species-level discrimination ability of partial COI within *Manouria* and *Indotestudo* genera.

Methods

The land tortoises inhabiting in the wild habitat of NE India were identified through previously reported morphological key characteristics [7,8,12,16-18]. Morphological characters along with photographs were recorded, and the required amount of blood samples were collected from the right hind limb of different individuals and subsequently stored in the buffer at 4°C. The investigators acquired permission to conduct the research under section 28 of the Wildlife (Protection) Act, 1972 (as amended up to date) from the Chief Conservator of Forest (Wildlife) Assam. The population of the target group of animals is very scarce in the nature, so this study relied solely on judicious sampling.

Genomic DNA was extracted from each of the blood samples using standard phenol-chloroform extraction method and subsequently precipitated with a double volume of chilled ethanol [31]. Thereafter, the extracted DNA was dissolved in 200µl nuclease-free water and stored at (-80)°C. The barcode segment was amplified using the published primer pair [32]. The 25 µl PCR reaction mixture contains 10 pmol of each primer, 25-50 ng of DNA template, 1x PCR buffer, 1.0–1.5 mM of MgCl₂, 0.25 mM of each dNTPs, and 0.25 U of high-fidelity *Taq*DNA polymerase (Applied Biosystems Inc., CA, USA). Amplification was performed using a Veriti® Thermal Cycler (Applied Biosystems Inc., CA, USA) and sequenced bi-directionally using automated DNA sequencer (ABI3500, Applied Biosystems Inc., CA, USA).

The sequences were screened and retrieved from each of the bidirectional sequences by using the Sequence Scanner Software v1.0. Applied Biosystem and ~2% overlap bases were discarded. The developed nucleotide sequences were checked by BLASTn searches and Bold-IDs, and correct amino acid sequences were determined by checking the open reading frames (ORF) using ORF prediction by online software and subsequently submitted to GenBank. Six self developed sequences and 12 database sequences were incorporated for the comparison of congeneric and conspecific divergences of the Indian tortoise taxa. All the sequences were aligned to form combined data set of 18 sequences using the ClustalX program [33]. In order to follow the recommended criteria of distance based phylogeny, the Kimura-2-Parameter (K2P) models for nucleotide substitutions of the studied species was evaluated using MEGA 5 [34,35]. Pairwise nucleotide divergences were calculated using the K2P model and the Neighbour- Joining (NJ) tree on ~627 bp was constructed using the defacto K2P model with 1000 replicates.

Results

The collected wild living individuals having the joined pectoral scute were taxonomically depicted as *Manouria emys phayrei*, a subspecies of *Manouria emys* (Figures 1D,E). The three individuals of *Indotestudo* from NE India (23.36 N 93.00 E) were identified as *Indotestudo elongata*, with the presence of nuchal scute and the ratio of interhumoral suture to interpectoral suture being 0.87 (Figures 1B,B1). One individual of *Indotestudo* collected from the same site appeared to be *Indotestudo forstenii*, with the absence of nuchal scute and the ratio of interhumoral suture being 1.66 (Figures 1C,C1).

Table 1 shows the comprehensive identification results based on GenBank and BOLD databases as well as the existing status of the Indian tortoise species. The developed sequences of Manouria emys showed 90% - 93% identity with their singlet congeners, Manouria impressa and 90% maximum identity with nearest neighbor (NN) genus Chelonoidis in both GenBank and BOLD databases. The genus Indotestudo and its two congeners remained inconclusive in both BLASTn and Bold-IDs identity results. The three developed sequences of Indotestudo elongata showed 96% - 99% identity with the first NN congeners and 94% - 97% with the second NN congeners in both databases. However, the acquired sequences of Indotestudo elongata from GenBank showed 96% - 99% identity with the first NN congeners in both databases but showed 97% identity with the second NN congeners only in Bold-IDs. Furthermore, the developed sequence of Indotestudo forstenii showed 96% - 97% identity with the first NN congeners and 97% with the second NN congeners in both databases. The acquired sequence of Indotestudo forstenii found 96% identical with the second NN congeners only in Bold-IDs but no match in GenBank. The two studied species of Indotestudo also showed 92% identity with nearest neighbor (NN) genus Chelonoidis in both GenBank and BOLD databases.

In the dataset, the interspecific K2P divergences were in the range of 1.43% \pm 0.004 to 15.75% \pm 0.017 within all the named Indian tortoise species (Table 2). The Indotestudo and Manouria genera showed 1.43% ± 0.004 to 5.31% ± 0.009 and 10.05% ± 0.013 congeneric divergence with their relevant respective congeners. Geochelone elegans, one extant species of India shows 12.92% ± 0.016 to 14.85% ± 0.017 and 15.24% ± 0.017 divergences from Indotestudo and Manouria genera respectively. The intraspecific K2P divergences were 1.86% ± 0.004 for Manouria emys, 0.44% ± 0.001 for Manouria impressa and 1.12% ± 0.002 for Indotestudo elongata yet, the intraspecific divergences were not calculated for Indotestudo travancorica, Indotestudo forstenii and NE Indian Indotestudo forstenii due to the lack of barcode sequences in databases as well as in the current study. The named Indian species Geochelone elegans, Manouria emys and its congeners Manouria impressa showed a minimum of 10.05 ± 0.013 interspecific divergences, that holds two fold gap with the interspecific divergences generated by the congeners of Indotestudo genus.

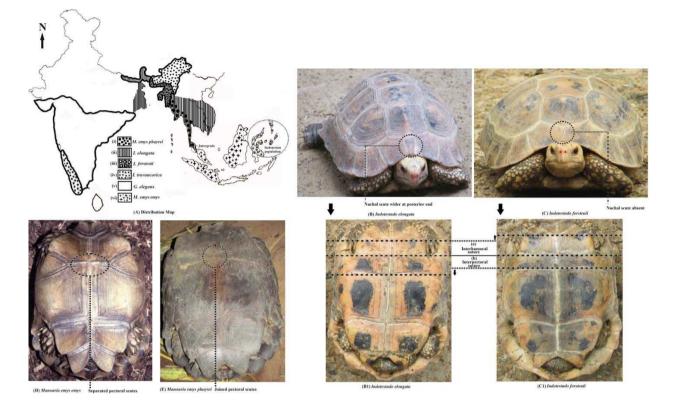


Figure 1. A distribution map of Manouria and Indotestudo genus, figure not to scale (A). Key character distinction between Indotestudo elongata (with Nuchal Scute) (B) and Indotestudo forstenii (without Nuchal Scute) (C). The ratio of Interhumoral to Interpectoral Suture is 0.87 in Indotestudo elongata (B1) and 1.66 in Indotestudo forstenii (C1). Morphological differences between M. emys emys having separated pectoral scutes (Photograph was collected from www.dght.de/ag_schildkroeten/ aktropland/ bilder/ 084.jpg) (D) and M. emys phayrei having joined pectoral scutes (E).

The two developed sequences of *Manouria emys* clustered separately from the GenBank *Manouria emys* sequences with 100 bootstrap supports. The NJ phylogeny readily discriminated the three studied genera of Indian tortoises into distinct clusters by their respective congeners and conspecies with significant bootstrap supports. However, both the developed and database sequences of *Indotestudo elongata*, *Indotestudo travancorica* and *Indotestudo forstenii* showed nonsynonomous clustering (Figure 2).

Discussion

The NE India region has been hitherto reportedly inhabited by two land tortoises viz., *Manouria emys* and *Indotestudo elongata*; while the Indonesian *Indotestudo forstenii* and south Indian *Indotestudo travancorica* were claimed both to be endemic through conventional taxonomy, supported also by molecular approaches [36]. Based on morphometric analysis, two distinct geographical populations of *Manouria emys* were given the rank of subspecies, viz., *Manouria emys phayrei* and *Manouria emys emys* (Figure 1D,E) [12]. The collected wild living individuals of *Manouria emys* from NE India exhibited morphological characters similar to those defined for the northern Asiatic subspecies *Manouria emys phayrei* (Figure 1E). BLASTn and Bold-IDs results for our query sequences found similarity with the complete mitochondrial sequences of Manouria emys (Accession No. NC007693 and DQ080040; Specimen Voucher MVZ238129 of the Museum of Vertebrate Zoology and California Academy of Sciences). Accordingly, the generated barcode sequences have been submitted to GenBank with the name Manouria emys. The developed barcode sequences of Manouria emys (n=2), however, exhibited no difference between themselves but exhibited 1.86% \pm 0.004 intraspecific K2P divergence with database sequences (n=2). Moreover, NJ tree with 100 bootstrap repetitions among the two data sets (developed and GenBank sequences) depicted a separate clade of Manouria emys. The observed genetic divergence between the two datasets of Manouria emys may perhaps relate to subspecies status as proposed previously through morphological studies [12]. Had the locality information of the database sequences of Manouria emys been given, it would have been easier to speculate about subspecies status with molecular evidence.

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Furthermore, the congeners of *Indotestudo* reportedly inhabit distant geographical locations; among the congeners, *Indotestudo forstenii* [11] is the oldest named species and is believed to be restricted in Sulawesi, Indonesia; *Indotestudo elongata* [37] is distributed through East and North East India, Myanmar, LAOs, Thailand, Cambodia to Vietnam; and *Indotestudo travancorica* [38] is restricted in South India. As stated above, *Indotestudo travancorica* is difficult to

Status NN Congeners1 NN Congeners2 NN Different genus (% identity) (% identity) (% identity) Common Name Scientific Name Accession CITES IWPA IUCN No. 100 100 85 6 95 6 95 6 GU563917 Asian forest tortoise or Asian brown tortoise Manouria emys JN794084 NC 007693 DQ080040 JX049141 Schedule IV Endangered Appendix II JN794083 Indotestudo elongata Eongated Tortoise JN794082 *NC_007695 *DQ656607 *DQ080043 Indotestudo forstenii Forsten's Tortoise *JX049140 -*NC_007696

Table 1. Common names, species status, scientific name, accession number and database comparison of the Northeast Indian tortoise species.

Asterisk (*) indicates the ambiguous identification for I. elongata and I. forstenii sequences from both developed and GenBank database. Blue bar indicates the BLASTn result and Gray bar indicates the Bold-IDs result. CITES=Convention on International Trade in Endangered Species of Wild Fauna and Flora, Appendix I= Most threatened species (No trade allowed), Appendix II=Species likely to be threatened due to unregulated trade (Trade is allowed through licensing). IWPA= Indian Wildlife Protection Act, 1972, IUCN=International Union for Conservation of Nature and Natural Resources, NN= Nearest Neighbor VERSITA

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Indian Tortoises	I. elongata	(NE) I.forstenii	I.travancorica	I.forstenii	M.emys	M.impressa	G.elegans
Indotestudo elongata	1.12 ± 0.002						
(NE) Indotestudo forstenii	2.57 ± 0.005						
Indotestudo travancorica	1.43 ± 0.004	2.69 ± 0.006	n/c				
Indotestudo forstenii	4.06 ± 0.007	5.31 ± 0.009	3.99 ± 0.008				
Manouria emys	13.77 ± 0.015	15.75 ± 0.017	14.25 ± 0.016	14.48 ± 0.017	1.86 ± 0.004		
Manouria impressa	14.39 ± 0.016	15.35 ± 0.017	14.75 ± 0.016	15.00 ± 0.017	10.05 ± 0.013	0.44 ± 0.001	
Geochelone elegans	14.02 ± 0.016	14.85 ± 0.017	14.71 ± 0.017	12.92 ± 0.016	15.24 ± 0.017	15.24 ± 0.017	n/c

Table 2. K2P divergence of Indian tortoise species. The bold letter indicates the intraspecific divergences within the studied species. n/c denoted for not calculated intraspecific data due to the lack sequences of the particular taxa in databases.

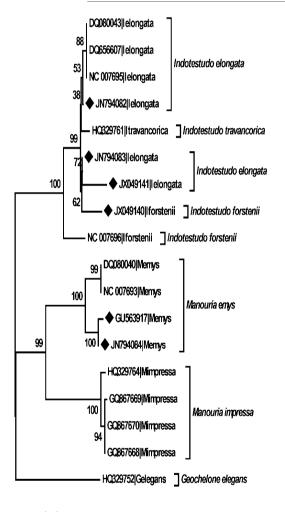




Figure 2. NJ Phylogeny of Indian tortoise species. Numbers along brunches indicate the bootstrap values. Black diamonds indicate the developed sequences for the current study.

distinguish, so it was once considered synonymise species of *Indotestudo forstenii* [39]. Further, the previous hypothesis that recognized Indonesian *Indotestudo forstenii* as being distinct from Indotestudo elongata appears intuitive. Although the wild living Indotestudo species are difficult to tell apart by morphological characteristics, the existing taxonomical characters supported the collected specimens as Indotestudo elongata (Figures 1B,B1) and Indotestudo forstenii (Figures 1C,C1), indicating that they inhabit the boundaries of Assam and Mizoram (23.36 N 93.00 E). However, morphological variations among the congeners of Indotestudo, predominantly in the length of interhumoral suture relative to interpectoral suture as observed in this study (Figures 1B1,C1), may relate to ecotypic variation among distant populations of the same species [40-43]. This study has given the morphological key evidences of Indotestudo forstenii, with existence in NE India eliminating the prevailing hypothesis of endemisms in Indonesia. In the study, it is further noted that the nuchal scute, an identifying character in Indotestudo elongata, does not remain always narrow but varies in width (posterior end wider than the anterior end) among different individuals at different life stages.

The perplexing species Indotestudo elongata. Indotestudo forstenii, and Indotestudo travancorica exhibited morphological differences, but displayed less distinctive genetic divergences at the species level in barcode sequences. The NJ phylogeny reflects nonsynonomous clustering by all the congeners of Indotestudo, which had the least interspecific divergences from the other species, and which violated the sequence divergence criteria for distinct species, as reported for several other turtles and tortoises [28]. It is evident in a few cases, however, that low rates of both molecular evolution and chromosomal rearrangement in turtles [42,45] may make hybridization possible by delaying the evolution of genetic barriers to reproduction. Further, because mitochondrial genes tend to accumulate differences at a rate several-fold slower in turtles than in other vertebrates, species considered 'recent radiations' will probably be nearly alike [28]. It is evident in this study that the congeners of Indotestudo are genetically very much identical at the COI partial gene segment. If, the replica sequences of Indotestudo congeners are enriched in the database from wide geographical ranges the synapomorphic character among the congeners would be substantiated and rectify the ambiguous species taxonomy.

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References

- Turtle Taxonomic Working Group (2007) An annotated list of modern turtle taxa with comments on areas of taxonomic instability and recent change. Chelonian Res Monogr 4: 173-199.
- [2] Ernst CH, Barbour RW (1989) Turtles of the World. Washington, DC: Smithsonian Institution Press.
- [3] IUCN (2011) IUCN Red List of Threatened Species. Version 2011.2. www.iucnredlist.org.
- [4] Gibbons JW, Scott DE, Ryan TJ (2000) The global decline of reptiles, deja vu amphibians. BioScience 50: 653-666.
- [5] Van Dijk PP, Stuart BL, Rhodin AGJ (2000) Asian Turtle Trade – Proceedings of a Workshop on Conservation and Trade of Freshwater Turtles and Tortoises in Asia. Chelonian Research Foundation, Lunenburg.
- [6] Turtle Conservation Fund (2002) A global action plan for conservation of tortoises and freshwater turtles, Strategy and funding prospectus, Conservation International and Chelonian Research Foundation, Washington, DC, pp 30.
- [7] Das I (1991) Colour guide to the turtles and tortoises of the Indian Subcontinent, R and A Publishing Limited, Portishead, U. K.
- [8] Das I (1995) Turtles and tortoises of India, Oxford University Press, Bombay, India.
- [9] Das KC, Gupta A (2011) Site records of softshell turtles (Chelonia: Trionychidae) from Barak Valley, Assam, northeastern India. Journal of Threatened Taxa 3: 1722-1726.
- [10] UNEP-WCMC (2012) UNEP-WCMC Species Database: CITES-Listed Species.
- [11] Schlegel H, Muller S (1840) Over de Schildpadden van den Indischen Archipel., en beschrijving eener nieuwe soort van Sumatra, In: Verhandelingen over de natuurlijke geschiendenis der Nederlandsche Overzeesche Bezittingen, Zoologie, Schildpadden. Lieden, 3: 29-36.
- [12] Schaffer C, Morgan V (2002) Behavioral Observations of Captive Juvenile Manouria emys phayrei with Notes on Degrees of Intergradation with Manouria emys emys. Turtle and Tortoise Newsl 5: 2-6.
- [13] Williams EE (1952) A new fossil tortoise from Mona Island, West Indies, and a tentative arrangement of the tortoises of the world. Bull Am Mus Nat Hist 99:541-560.
- [14] Crumly CR (1982). A cladistic analysis of Geochelone using cranial osteology. J Herpetol 16: 215-234.
- [15] Crumly CR (1984). A hypothesis for the relationship of land tortoise genera (family Testudinidae). Studia Geologica Salmanticensia, Studia Palaeochelonologica 1:115-124.

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- [16] Pritchard PCH (2000) Indotestudo travancorica...Avalid species of tortoise? Reptile & Amphibian Hobbyist 5: 18-28.
- [17] Ives I (2006) Conservation of Sulawesi's two endemic chelonians, Leucocephalon yuwonoi and Indotestudo forstenii; a preliminary investigation into in-situ and ex-situ conservation concerns. Thesis, Antioch University New England.
- [18] Ives I, Spinks PQ, Shaffer HB (2008) Morphological and genetic variation in the endangered Sulawesi tortoise Indotestudo forstenii: evidence of distinct lineages? Conserv Genet 9:709-713.
- [19] Stoeckle M (2003) Taxonomy, DNA, and the bar code of life. BioScience 53: 2-3.
- [20] Bury RB (1979) Population ecology of freshwater turtle. Turtle's prospectives and research. John Wiley & Sons. New york, 571- 602.
- [21] Pritchard PCH (1979) Encyclopedia of Turtles. T.F.H. Publications, Neptune, N.J., 895.
- [22] Moll EO (1984) India's freshwater turtles in India their status, conservation and management. Hamadryad 9: 49-55.
- [23] Ratnasingham S, Hebert PDN (2007) BOLD : The Barcode of Life Data System . Mol Ecol Notes 7: 355–364.
- [24] Hebert PDN, Ratnasingham S, deWaard JR (2003) Barcoding animal life: cytochrome c oxidase subunit 1 divergences among closely related species. Proc R Soc Lond B Biol Sci 270: S96–S99.
- [25] Hebert PDN, Stoeckle MY, Zemlak TS, Frances CM (2004) Identification of birds through DNA barcodes. PLoS Biol, 2: 1657-1663.
- [26] Vargas SM, Araujo FC, Santos FR (2009) DNA barcoding of brazilian sea turtles (testudines). Genet Mol Biol 32: 608-612.
- [27] Naro-Maciel E, Reid B, Fitzsimmons NN, Le M, Desalle R, Amato G (2010) DNA barcodes for globally threatened marine turtles: a novel registry approach to documenting biodiversity. Mol Ecol Resour 10: 252-263.
- [28] Reid BN, Le M, McCord WP, Iverson JB, Georges A, Bergmann T, Amato G, Desalle R, Naro-Maciel E (2011) Comparing and combining distance-based and characterbased approaches for barcoding turtles. Mol Ecol Resour 11: 956-967.
- [29] DeSalle R, Egan MG, Siddall M (2005) The unholy trinity: taxonomy, species delimitation and DNA barcoding. Philos Trans R Soc Lond B 360: 1905-1916.
- [30] Kelly RP, Sarkar IN, Eernisse DJ, Desalle R (2007) DNA barcoding using chitons (genus Mopalia). Mol Ecol Notes 7: 177-83.



- [31] Sambrook J, Russell DW (2001) Molecular cloning a laboratory manual, (Cold Spring Harbor Laboratory Press, Cold Spring Harbor).
- [32] Ward RD, Zemlak TS, Innes BH, Last PR, Hebert PDN (2005) DNA barcoding Australia's fish species. Philos Trans R Soc Lond B 360: 1847-1857.
- [33] Thompson JD, Gibson TJ, Higgins DG (2002) Multiple sequence alignment using ClustalW and ClustalX. Curr Protoc Bioinformatics 2: 23.
- [34] Kimura M (1980) A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. J Mol Evol 16:111-120.
- [35] Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S (2011) MEGA5: Molecular Evolutionary Genetics Analysis using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods. Mol Biol Evol 28: 2731-2739.
- [36] Iverson JB, Spinks PQ, Shaffer HB, McCord WP, Das I (2001)
 Phylogenetic relationships among the Asian tortoises of the genus Indotestudo (Reptilia: Testudines: Testudinidae).
 Hamadryad 26: 272 - 275.
- [37] Blyth E (1853) Notices and descriptions of various reptiles, new or little-known. J. Asiat. Soc. Bengal 22: 639-655.
- [38] Boulenger GA (1907) A new tortoise from Travancore. J Bombay Nat Hist Soc 17: 560-564.

- [39] Hoogmoed MS, Crumly CR (1984) Land tortoise types in the Rijksmuseum van Natuurlijke Histoire with comments on nomenclature and systematics (Reptilia: Testudines: Testudinidae). Proc Zool Soc Lond 58: 241-259.
- [40] Hausmann A, Haszprunar G, Hebert PD (2011) DNA barcoding the geometrid fauna of Bavaria (Lepidoptera): successes, surprises, and questions. PLoS ONE 6: 223-24.
- [41] Park DS, Foottit R, Maw E, Hebert PD (2011) Barcoding bugs: DNA-based identification of the true bugs (Insecta: Hemiptera: Heteroptera). PLoS ONE 6: e18749.
- [42] Laskar BA, Bhattacharjee MJ, Dhar B, Mahadani P, Kundu S, et al. (2013) The Species Dilemma of Northeast Indian Mahseer (Actinopterygii: Cyprinidae): DNA Barcoding in Clarifying the Riddle. PLoS ONE 8: e53704.
- [43] Bhattacharjee MJ, Laskar BA, Dhar B, Ghosh SK (2012) Identification and Re-Evaluation of Freshwater Catfishes through DNA Barcoding. PLoS ONE 7: e49950.
- [44] Bickham JW (1981) Two-hundred-million-year-old chromosomes: deceleration in the rate of karyotypic evolution in turtles. Science 212: 1291-1293.
- [45] Avise JC, Bowen BW, Lamb T, Meylan AB, Bermingham E (1992) Mitochondrial DNA evolution at a turtle's pace: evidence for low genetic variability and reduced microevolutionary rate in Testudines. Mol Biol Evol 9: 457- 473.